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IN VITRO ANTIBACTERIAL ACTIVITY OF METHANOLIC EXTRACT OF WILD MUSHROOMS FROM SOUTHERN WESTERN GHATS, INDIA

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ABSTRACT

The present study is aims to investigate the antibacterial activities of crude methanol extracts of 24 southern Western Ghats wild mushrooms. Crude methanol extracts from 24 mushrooms from southern Western Ghats, India were evaluated for their antibacterial activity against *Eggerthella lenta*, *Vibrio parahaemolyticus* and *Enterococcus faecalis* by agar well diffusion method. The study revealed that about 67% of the mushrooms inhibited growth of all the test bacteria, 29% was active against any of the two test bacteria and 4% were completely inactive. Amoxicillin used as standard reference. 15 mushroom species showed strong antibacterial activity against *Eggerthella lenta* higher than the standard antibiotic (Amoxicillin). 11 mushrooms showed strong antibacterial activity against *Vibrio parahaemolyticus* higher than the standard antibiotic (Amoxicillin). The best *in vitro* antibacterial activity was by *Gymnopilus junonius* (26.0 mm against *Eggerthella lenta*; 25.0 mm against *Vibrio parahaemolyticus*) followed by *Tricholoma equestre* (21.0 mm against *Vibrio parahaemolyticus*). *Gymnopilus junonius, Tricholoma equestre* and *Trametes versicolor* have higher antibacterial activity than that of standard antibiotic.

Keywords: Antibacterial activity, Macrofungi, Western Ghats, Gymnopilus junonius, Tricholoma equestre, Trametes versicolor.

INTRODUCTION

Mushrooms are a promising source for a variety of potential antimicrobial compounds and are relatively less explored. They have been shown to be rich sources of natural antibiotics [1] and accumulate a variety of chemicals with strong anti-oxidant properties [2]. The search for new natural antimicrobial agents which have no or low impact on the environment as well as human health are highly needed in the present scenario of development of drug resistance in pathogenic bacteria, emergence of new microbial diseases and adverse side effects of synthetic drugs. Wild and cultivated mushrooms contain a huge diversity of biomolecules with nutritional [3] and/or medicinal properties [4-7]. Numerous mushroom extracts have been reported as having antimicrobial activity against bacteria.

As a matter of fact, macrofungi need secondary metabolites for their survival in various harsh environmental conditions. These chemical compounds are usually antibacterial and antifungal in nature. Therefore, antimicrobial compounds could be isolated from many mushroom species and could be of benefit for humans.

Bioactive molecules have been isolated not only from edible, but also from inedible species [8-9]. The mushrooms and their medicinal properties, have long been recognised in China, Korea, and Japan, e.g., hypotensive and renal effects [10-11], immunomodulatory and antitumour activities of polysaccharide-protein complex (PSPC) from mycelial cultures, immunomodulatory and antitumour activities of lectins from edible mushrooms [12-18], isolation and characterization of a Type-I Ribosome-Inactivation protein from Volvariella volvacea [19], and medicinal effects of Ganoderma lucidum [20-21]. The responsible bioactive compounds belong to several chemical groups which are often polysaccharides or triterpenes [22-23]. One macrofungi species can have various bioactive compounds and pharmacological effects [5].

A significant amount of work has been carried out on the antimicrobial activities and chemical content of medicinal mushrooms including edible one but in Western Ghats of Tamil Nadu has not yet been explored for their antibacterial potential. Therefore, this study seeks to

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identify and survey the properties of the species of wild mushrooms growing in Western Ghats of Tamil Nadu state in India.

MATERIALS AND METHODS Collection and preparation of mushroom species

Macrofungal species were collected from November 2013 to October 2014 from from southern Western Ghats in the Fingerpost Reserve Forest of North Zone of Nilgiri, Tamilnadu, India. In order to have wide range of species as possible, specimens were collected from forests, fields and woods. Identification of the mushrooms was done by comparing their morphological, anatomical and physiological traits with the standard description and the keys provided by the Directorate of Mushroom Research, Solan, India. The mushrooms were dried in the shade to prevent mushroom cells from sun light which destroy the cell and ground to powder using mortar and pestle.

Extracts preparation

To perform antimicrobial activity, fruiting bodies of the mushroom samples air-dried at 40°C and300mg were ground using the pestle and mortar with methanol and then filtered using WhatmanNo.1 filter paper. After that, the extract was centrifuged at 5000rpm for 15min and the supernatant was stored at 4°C for further experiment.

Antimicrobial activity

The antimicrobial activities of each of the methanolic extracts were tested against standard Gram positive bacteria (*Eggerthella lenta* (ATCC 43055), *Enterococcus faecalis* (ATCC 29212) and Gram negative bacterium (*Vibrio parahaemolyticus* (MTCC 451). Kirby-Bauer well diffusion assay technique [24-25] was used here. Muller-Hinton agar served as the basal medium to carry out the assay. Sterilized media plates were seeded with bacterial suspension using sterile swab. Wells (6mm) were loaded with fungal extracts at a desired concentration and were placed onto the bacteria seeded plates. Methanol was used as negative control and antibiotic as positive control. The plates were incubated at 37°C for 24 h. Two

replicates were performed for each extract against each of the tested organisms. Simultaneously, controls involving the addition of amoxicillin and methanol instead of the extracts were included. Upon the completion of incubation the diameter of the resultant inhibition zones were measured and tabulated as means.

RESULTS

In the present experiment, 24 species of wild mushroom collected from the Western Ghats of Tamil Nadu, India were evaluated for their antibacterial potential. All the mushrooms used in this study were found to exhibit various degrees of antimicrobial effects against the tested microorganisms (Plate 1, 2 & 3). The zone of inhibition exhibited more than the positive control *i.e.*, Amoxicillin was considered as highly active for extracts. The study revealed that about 67% of the mushrooms inhibited growth of all the test bacteria, 29% was active against any of the two test bacteria and 4% were completely inactive (Table 1). The best in vitro antibacterial activity was by Gymnopilus junonius (26.0 mm against Eggerthella lenta; 25.0 mm against Vibrio parahaemolyticus) followed by Tricholoma equestre (21.0 mm against Vibrio parahaemolyticus).

15 mushroom species showed strong antibacterial activity against *Eggerthella lenta* higher than the standard antibiotic (Amoxicillin). 11 mushrooms showed strong antibacterial activity against *Vibrio parahaemolyticus* higher than the standard antibiotic (Amoxicillin). Whereas, only 4 mushroom species have showed strong antibacterial activity against *Enterococcus faecalis* higher than the standard antibiotic (Amoxicillin).

Figure 1 summarizes the antibacterial activity of methanolic extract of various wild mushrooms of Western Ghats with Positive control in relation to their susceptibility to the tested microorganisms. These results reveal that *Gymnopilus junonius, Tricholoma equestre* and *Trametes versicolor* had comparatively similar concentrations of standard antibiotics, which confirms the presence of bioactive components in these wild mushrooms.









Table 1. Antibacterial activity of wild mushrooms from south Western Ghats

Sample No	Mushroom Species	Zone of Inhibition in mm for 100 µg/mL of crude extract			
		Gram Positive		Gram Negative	
		Enterococcus faecalis	Eggerthella lenta	Vibrio parahaemolyticus	
1	Trametes versicolor	15	16	18	
2	Calvatia gigantea	13	12	16	
4	Lactarius sp.	-	-	-	
6	Schizophyllum commune	7	5	-	
7	<i>Clitocybe</i> sp.	9	8	-	
8	Gymnopilus junonius	17	26	25	
9	Lycoperdon pyriforme	-	-	8	
10	Cortinarius sp.	11	13	13	
12	Collybia radicata	10	10	11	
13	<i>Hygrocybe</i> sp.	9	9	13	
15	Tricholoma sp.	12	14	10	
16	Unknown sp.	9	14	11	

19	Unknown sp.	7	14	12
20	Russula olivacea	8	16	11
21	Unknown sp.	11	18	-
22	Unknown sp.	12	9	6
25	<i>Mycena</i> sp.	13	15	14
26	Unknown sp.	7	14	-
27	Coprinellus disseminatus	12	18	-
28	Daldinia concentrica	-	7	9
30	Coprinus comatus	12	16	14
31	Unknown sp.	14	15	16
32	Amanita muscaria	10	13	13
35	Tricholoma equestre	17	17	21
	Amoxicillin	14	13	12
	Control	-	-	-

(-) = NZI - No Zone of Inhibition/Absence of susceptibility

DISCUSSION

Methanolic extract of Agaricus bisporus, the most cultivated mushroom in the world, showed activity against Bacillus cereus, Micrococcus luteus, Micrococcus flavus, Staphylococcus aureus, and Staphylococcus epidermidis [26-28]. Several other Agaricus species have also demonstrated antimicrobial activity. Methanolic extracts Agaricus bitorquis and Agaricus essettei showed an inhibitory effect upon all the tested gram-positive bacteria [28]. Methanolic extract of Agaricus silvicola. Cantharellus cibarius, Lentinus edodes, Boletus edulis and different Cortinarius sp. also revealed antimicrobial properties against Staphylococcus aureus and in some cases Bacillus cereus and Bacillus subtilis [29-34]. The mycelium of Agaricus cf. nigrecentulus and Tyromyces

duracinus (ethyl acetate extracts) showed activity only against *Staphylococcus saprophyticus* [35].

Eggerthella lenta is an emerging pathogen which is susceptible to amoxicillin-clavulanate, cefoxitin, metronidazole, ertapenem, piperacillin-tazobactam, and meropenem; resistant to penicillin and piperacillintazobactam [36]. The present study is, the first of its kind, finding the antibacterial activity by crude extract of wild mushrooms against *Eggerthella lenta*.

Lactarius sp. showed no activity against all the tested bacteria. However, Aqueous and organic (hexane, ethyl acetate and methanol) basidiocarp extracts of the edible mushroom Lactarius indigo were showed varied activity against various diarrheagenic Escherichia coli strains, Pseudomonas aeruginosa, Enterobacter cloacae, Staphylococcus aureus and Salmonella enterica [37].

The extracts obtained from six Lactarius species such as L. deterrimus, L. sanguifluus, L. semisanguifluus, L. piperatus, L. deliciosus and L. salmonicolor showed varied inhibiting activity against Escherichia coli, Micrococcus luteus, Stapylococcus aureus, Salmonella thyphi, Klebsiella pneumoniae, Pseudomonas aeruginosa, Corynebacterium xerosis, Bacillus cereus, Bacillus megaterium, Mycobacterium smegmatis, Candida albicans but no antagonistic effect on Saccharomyces cerevisiae [38].

Lactarius controversus showed no activity against Candida albicans, Escherichia coli, Pseudomonas aeruginosa, Salmonella enterica and showed activity against Shigella flexneri as well as at higher concentration $(30 \Box L)$ showed inhibiting activity against Pseudomonas aeruginosa whereas Lactarius deliciosus showed no activity against Candida albicans, Escherichia coli, Salmonella enterica and showed inhibiting activity against Pseudomonas aeruginosa and Shigella flexneri [39].

The antimicrobial activity of aqueous, methanol, hexane, and ethyl acetate extracts from edible wild and cultivated mushrooms such as *Agaricus silvicola*, *Clitocybe nebularis* and *Tricholoma equestre* against *Vibrio parahaemolyticus* and *Staphylococcus aureus*, foodborne pathogenic bacterial strains was screened and antimicrobial activity of gram-positive bacteria were more sensitive than gram-negative bacteria to fungal extracts [40].

Gymnopilus junonius a poisonous mushroom, formerly known as Gymnopilus spectabilis, belonging to the family Cortinariaceae, is found growing in dense clusters on stumps and logs of hardwoods and conifers. Two polyacetylenes were isolated from Gymnopilus spectabilis, namely hepta-4,6-diyn-3-ol and 7-chlorohepta-4,6-diyn-3-ol which showed biological activity against Bacillus brevis, B. subtilis, Streptococcus pyogenes and Staphylococcus aureus [41]. Extracts of Gymnopilus spectabilis showed positive antibacterial activity against Staphylococcus aureus, Bacillus subtilis and resistance to Escherichia coli [42]. A lectin was isolated from fruiting bodies of the mushroom Gymnopilus spectabilis inhibited the growth of Staphylococcus aureus and Aspergillus niger [43].

Polysaccharides, sesquiterpenes, lectins, phenolic compounds, α -d-glucan and other biomolecules with different therapeutic effects (antibacterial, antifungal, cytotoxic, antitumor, anti-inflammatory, insecticidal, nematocidal, antioxidant and others) were detected in several medicinal ectomycorrhyzae forming basidiomycetous mushrooms esp. various species of *Tricholoma* such as *T. lobayense*, *T. giganteum*, *T. matsutake*, *T. mongolicum* and *T. portentosum* [44-46].

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CONFLICT OF INTEREST No interest

REFERENCES

- 1. Kupra J, Anke T, Oberwinkler F, Schramn G, Steglich W. Antibiotics from basidiomycetes. VII. Crinipellin, a new antibiotic from the basidiomycetous fungus *Crinipellis stipitaria* (Fr.) *Pat. J. Antibiot*, 32, 1979, 130-135.
- 2. Sun T, Tang J, Powers JR. Antioxidant activity and quality of asparagus affected by microwave-circulated water combination and conventional sterilization. *Food Chem*, 100, 2007, 813-819.
- 3. Kalac P. Chemical composition and nutritional value of European species of wild growing mushrooms: A review. *Food Chem*, 113, 2009, 9-16.
- 4. Borchers A, Keen CL, Gershwin ME. Mushrooms, tumors, and immunity: an update. Exp. Biol. Med, 229, 2004, 393-406.
- 5. Lindequist U, Niedermeyer THJ, Jülich WD. The pharmacological potential of mushrooms. CAM, 2, 2005, 285-299.
- 6. Poucheret P, Fons F, Rapior S. Biological and pharmacological activity of higher fungi: 20-Year retrospective analysis. *Mycologie*, 27, 2006, 311-333.
- 7. Alves MJ, Ferreira ICFR, Dias J, Teixeira V, Martins A, Pintado M. A Review on Antimicrobial Activity of Mushroom (Basidiomycetes) Extracts and Isolated Compounds. *Planta Med*, 78(16), 2012, 1707-1718.
- 8. Quang DN, Hashimoto T, Asakawa Y. Inedible mushrooms: A good source of biologically active substances. *Chem. Record*, 6, 2006, 79-99.
- 9. Nedelkoska DN, Pančevska NA, Amedi H, Veleska D, Ivanova E, Karadelev M. *et al.*, Screening of antibacterial and antifungal activities of selected Macedonian wild mushrooms. *Jour. Nat. Sci*, 124, 2013, 333-340.
- 10. Tam SC, Yip KP, Fung KP, Chang ST. Hypotensive and renal effects of an extract of the edible mushroom, *Pleurotus* sajor-caju. Life Sci, 38, 1986, 1155-1161.
- 11. Yip KP, Fung KP, Chang ST, Tam SC. Purification and mechanism of the hypotensive action of an extract from edible mushroom *Pleurotus sajor-caju. Neurosci. Lett. Suppl*, 28, 1987, S59.
- 12. Liu F, Ooi VEC, Chang ST. Antitumour components of the culture filtrates from *Tricholoma* sp. World J. Microbiol. *Biotech*, 11, 1995, 486-490.
- 13. Liu F, Ooi VEC, Liu WK, Chang ST. Immunomodulatory and antitumour activities of polysaccharide-protein complex from the culture filtrates of a local edible mushroom, *Tricholoma lobayense*. Gen. Pharmac, 27, 1996, 621-624.
- 14. Liu F, Fung MC, Ooi VEC, Chang ST. Induction in the mouse Gene expression of immunomodulating cytokines by mushroom polysaccharide-protein complexes. *Life Sci*, 58, 1996, 1795-1803.
- 15. Wang HX, Ng TB, Liu WK, Ooi VEC, Chang ST. Isolation and characterization of two distinct lectins with antiproliferative activity from the cultured mycelium of the edible mushroom *Tricholoma mongolicum*. *Int. J. Peptide Protein Res*, 46, 1995, 508-513.
- Wang HX, Ng TB, Ooi VEC, Liu WK, Chang ST. A polysaccharide peptide complex from cultured mycelia of the mushroom *Tricholoma mongolicum* with immune enhancing and antitumour activities. *Biochem. Cell Bio*, 74, 1996, 95-100.
- 17. Wang HX, Ng TB, Liu WK, Ooi VEC, Chang ST. Polysaccharide-peptide complexes from the cultured mycelia of the mushroom Coriolus versicolor and their culture medium activate mouse lymphocytes and macrophages. *Int. J. Biochem. Cell Bio*, 28, 1996, 601-607.
- 18. Wang HX, Ng TB, Ooi VEC, Liu WK, Chang ST. Actions of lectins from the mushroom *Tricholoma mongolicum* on macrophages, splenocytes and life-span in sarcombearing mice. *Anticancer Res*, 17, 1997, 419-424.
- 19. Yao QZ, Yu MM, Ooi LSM, Ng TB, Chang ST, Sun SSM. *et al.*, 1998. Isolation and characterization of a Type I ribosome-inactivating protein from fruiting bodies of the edible mushroom (*Volvariella volvacea*). *J. Agri. Food Chem*, 46, 1998, 788-792.
- 20. Chang ST, Buswell JA. *Ganoderma lucidum* (Curt.:Fr.) P. Karst. (Aphyllophoromycetidease): a mushrooming medicinal mushroom. *Int. J. Med. Mush*, 1, 1999, 139-146.
- Chang ST, Miles PG. Mushrooms: Cultivation, Nutritional Value, Medicinal Effect, and Environmental Impact. 2nd ed. CRC Press, Boca Raton, 2004.
- 22. Kim DM, Pyun CW, Ko HG, Park WM. Isolation of antimicrobial substances from *Hericium erinaceum*. *Mycobiology*, 28, 2000, 33-38.
- 23. Sun Y, Liu J. Purification, structure and immunobiological activity of a watersoluble polysaccharide from the fruiting body of *Pleurotus ostreatus*. *Bioresour*. *Technol*, 100, 2009, 983-986.
- 24. James JB. Antimicrobial susceptibility testing by the Kirby-Bauer disc diffusion method. Ann. *Clin. Lab. Sci*, 3 (2), 1973, 135-140.
- 25. NCCLS. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically: Approved standard. 5th ed. M7-A5, National Committee for Clinical Laboratory Standards, Wayne, Pennsylvania. 2000.
- 26. Tambekar DH, Sonar TP, Khodke MV, Khante BS. The novel antibacterials from two edible mushrooms: Agaricus bisporus and Pleurotus sajor-caju. Int. J. Pharmacol, 2, 2006, 584-587.
- 27. Ozen T, Darcan C, Aktop O, Turkekul I. Screening of antioxidant, antimicrobial activities and chemical contents of edible mushrooms wildly grown in the Black Sea region of Turkey. Comb. *Chem. High Throughput Screen*, 14 (2), 2011, 72-84.
- Ozturk M, Duru ME, Kivrak S, Mercan-Doğan N, Türkoglu A. In vitro antioxidant, anticholinesterase and antimicrobial activity studies on three Agaricus species with fatty acid compositions and iron contents: a comparative study on the three most edible mushrooms. *Food Chem. Toxic*, 49, 2011, 1353-1360.

- 29. Hirasawa M, Shouji N, Neta T, Fukushima K, Takada K. Three kinds of antibacterial substances from *Lentinus edodes* (Berk.) Sing. (Shiitake, an edible mushroom). *Intl. J. Antimicrob. Agents*, 11, 1999, 151-157.
- 30. Ishikawa NK, Kasuya MCM, Vanetti MCD. 2001. Antibacterial activity of *Lentinula edodes* grown in liquid medium. *Braz. J. Microbiol*, 32, 2011, 206-210.
- 31. Hur JM, Yang CH, Han SH, Lee SH, You YO, Park JC. et al., 2004. Antibacterial effect of *Phellinus linteus* against methicillin-resistant *Staphylococcus aureus*. *Fitoterapia*, 75, 2004, 603-605.
- 32. Barros L, Cruz T, Baptista P, Estevinho LM, Ferreira ICFR. Wild and commercial mushrooms as source of nutrients and nutraceuticals. *Food Chem. Toxicol*, 46 (8), 2008, 2742-2747.
- 33. Barros L, Cruz T, Baptista P, Estevinho LM, Ferreira ICFR. 2008a. Wild and commercial mushrooms as source of nutrients and nutraceuticals. *Food Chem. Toxicol*, 46, 2008, 2742-2747.
- 34. Barros L, Venturini BA, Baptista P, Estevinho LM, Ferreira ICFR. Chemical composition and biological properties of Portuguese wild mushrooms: a comprehensive study. J. Agric. *Food Chem*, 56, 2008, 3856-3862.
- 35. Rosa LH, Machado KMG, Jacob CC, Capelari M, Rosa CA, Zani CL. Screening of Brazilian Basidiomycetes for antimicrobial activity. *Mem. Inst. Oswaldo Cruz*, 98, 2003, 967-974.
- 36. Gardiner BJ, Tai AY, Kotsanas D, Francis MJ, Roberts SA, Ballard SA. *et al.*, Clinical and Microbiological Characteristics of *Eggerthella lenta* Bacteremia. J. Clin. Microbiol., 53(2), 2015, 626-635.
- 37. Ochoa A. Antibacterial and cytotoxic activity from basidiocarp extracts of the edible mushroom *Lactarius indigo* (Schw.) Fr. (Russulaceae). *Afr. J. Pharm. Pharmacol*, 5(2), 2011, 281-288.
- 38. Dulger B, Yilmaz F, Gucin F. Antimicrobial Activity of Some Lactarius Species. Pharm. Biol, 40(4), 2002, 304-306.
- 39. Altuner EM, Akata I. Antimicrobial activity of some macrofungi extracts. SAÜ. Fen Bilimleri Dergisi, 14(1), 2010, 45-49.
- 40. Venturini ME, Rivera CS, Gonzalez C, Blanco D. Antimicrobial activity of extracts of edible wild and cultivated mushrooms against foodborne bacterial strains. J. Food Prot, 71(8), 2008, 1701-1706.
- 41. Aqueveque P, Becerra J, Palfner G, Silva M, Alarcón J, Anke T. *et al.* Antimicrobial activity of metabolites from mycelial cultures of chilean basidiomycetes. *J. Chil. Chi. Soc*, 51(4), 2006, 1057-1060.
- 42. Garrido N, Becerra J, Marticorena C, Oehrens E, Silva M, Horak E. Antibiotic properties of ectomycorrhizae and saprophytic fungi growing on *Pinus radiata* D. *Don I. Mycopathologia*, 77(2), 1982, 93-98.
- 43. Alborés S, Mora P, Bustamante MJ, Cerdeiras MP, Fraguas LF. Purification and Applications of a Lectin from the Mushroom *Gymnopilus spectabilis*. *Appl. Biotechnol. Biochem*, 172 (4):, 2014, 2081-2090.
- 44. Badalyan SM. The main groups of therapeutic compounds of medicinal mushrooms. Med. Mycol, 3, 2001, 16-23.
- 45. Badalyan SM. Potential of mushroom bioactive molecules to develop healthcare biotech products. In: Proceedings of the 8th International Conference on Mushroom Biology and Mushroom Products, 2014, 373-378.
- 46. Mattila P, Könkö K, Eurola M, Pihlava JM, Astola J, Vahteristo L. *et al.*, Contents of vitamins, mineral elements, and some phenolic compounds in cultivated mushrooms. *J. Agric. Food Chem*, 49, 2001, 2343-2348.